

AD \_\_\_\_\_

Award Number: W81XWH-FE~~FE~~ Fİ

TITLE: OAT [ | ^ & | æ Á œ ã Á Ø & [ ~ } c ^ â Á | Á @ Á æ ð } æ ó ^ æ | ^ • Á - Á | ^ æ ó Ô æ & ^ | Á ^ || •

PRINCIPAL INVESTIGATOR: Öi EY ã æ \* ç Á ~

CONTRACTING ORGANIZATION: University of T ã æ ã  
T ã æ ã OSÁ-HFÎ Á

REPORT DATE: U& q à ^ | Á GFF

TYPE OF REPORT: OË } ~ æ

PREPARED FOR: U.S. Army Medical Research and Materiel Command  
Fort Detrick, Maryland 21702-5012

DISTRIBUTION STATEMENT: Approved for public release; distribution unlimited

The views, opinions and/or findings contained in this report are those of the author(s) and should not be construed as an official Department of the Army position, policy or decision unless so designated by other documentation.

REPORT DOCUMENTATION PAGE				Form Approved OMB No. 0704-0188	
Public reporting burden for this collection of information is estimated to average 1 hour per response, including the time for reviewing instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing this collection of information. Send comments regarding this burden estimate or any other aspect of this collection of information, including suggestions for reducing this burden to Department of Defense, Washington Headquarters Services, Directorate for Information Operations and Reports (0704-0188), 1215 Jefferson Davis Highway, Suite 1204, Arlington, VA 22202-4302. Respondents should be aware that notwithstanding any other provision of law, no person shall be subject to any penalty for failing to comply with a collection of information if it does not display a currently valid OMB control number. <b>PLEASE DO NOT RETURN YOUR FORM TO THE ABOVE ADDRESS.</b>					
1. REPORT DATE (DD-MM-YYYY) 01-10-2011		2. REPORT TYPE Annual		3. DATES COVERED (From - To) 15 Sep 2010 - 14 Sep 2011	
4. TITLE AND SUBTITLE A Molecular Basis Accounted for the Malignant Features of Breast Cancer Cells				5a. CONTRACT NUMBER	
				5b. GRANT NUMBER W81XWH-10-1-0417	
				5c. PROGRAM ELEMENT NUMBER	
6. AUTHOR(S) Dr. Xiangxi Xu  E-Mail: xxu2@med.miami.edu				5d. PROJECT NUMBER	
				5e. TASK NUMBER	
				5f. WORK UNIT NUMBER	
7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES) University of Miami Miami, FL 33136				8. PERFORMING ORGANIZATION REPORT NUMBER	
9. SPONSORING / MONITORING AGENCY NAME(S) AND ADDRESS(ES) U.S. Army Medical Research and Materiel Command Fort Detrick, Maryland 21702-5012				10. SPONSOR/MONITOR'S ACRONYM(S)	
				11. SPONSOR/MONITOR'S REPORT NUMBER(S)	
12. DISTRIBUTION / AVAILABILITY STATEMENT Approved for public release; distribution unlimited					
13. SUPPLEMENTARY NOTES					
14. ABSTRACT Malignancy is defined as the elevated mobility and invasiveness of tumor cells, and a deformed nuclear morphology is a common feature of malignant cells. We hypothesized that the decrease or absence of nesprin-1 in breast cancer cells may account for the malignant features of the neoplastic cells, the deformed nuclear morphology and invasiveness/high motility. We propose a pilot study to test this hypothesis. Indeed, we found that nesprin-1 expression is commonly lost in malignant breast cancer cell lines (Aim 1). We found that the suppression of nesprin-1 by siRNA led to nuclear morphological deformation and increased invasion (Aim 2). We also tested restoration of nesprin-1 expression in malignant breast cancer cells and nesprin-1 was not sufficiently stable to produce other significant phenotypes in the transfected cells, likely due to technical limitation (Aim 3). The results of these pilot experiments support the initial hypothesis of nesprin-1 as a metastatic suppressor gene and as an underlying link between two prominent features of a malignant cell, nuclear deformation and cellular malleability. We also realize further complexity of nesprin-1 function in breast cancer suppression. The pilot study promotes us to seek further investigation into the role of nesprin-1 in cancer malignancy.					
15. SUBJECT TERMS nuclear envelope, nesprin/SYNE, nuclear morphology, nuclear deformation, cellular malleability metastasis					
16. SECURITY CLASSIFICATION OF:			17. LIMITATION OF ABSTRACT  UU	18. NUMBER OF PAGES  8	19a. NAME OF RESPONSIBLE PERSON USAMRMC
a. REPORT U	b. ABSTRACT U	c. THIS PAGE U			19b. TELEPHONE NUMBER (include area code)

## Table of Contents

	<u>Page</u>
Introduction.....	4
Body.....	4
Key Research Accomplishments.....	7
Reportable Outcomes.....	7
Conclusion.....	8
References.....	8
Appendices.....	8

## INTRODUCTION:

The severity of cancer, known as the degree of malignancy, is caused by the ability of the cancer cells to move and invade tissues (malleability). Unlike normal cells which have a smooth and oval shape nucleus, the cancer cells often have deformed, irregular nuclear shape, and this phenomenon is used to diagnose the degree of malignancy, known as nuclear grade (1,2). In this project, we investigated an idea, that the two features of cancer cells, deformed nuclear shape and high mobility, may be caused by the loss of nesprin-1, a structural protein linking nuclear envelope to the cytoskeleton of cells (3-5). The pilot experiments are to study nesprin-1/Syne-1 as a metastatic/invasion suppressor gene and as an underlying link between two prominent features of a malignant cell, nuclear deformation and cellular malleability.

## BODY:

In the last year, we have followed the research plan described in the Statement of Work (SOW). We have accomplished most of the experiments proposed and reached the conclusion to support our hypothesis that loss of nuclear-cytoplasmic linker protein nesprin-1 in breast cancer cells may account for the malignant features of the neoplastic cells, the deformed nuclear morphology and invasiveness/high motility, and the loss of nesprin-1 may account for the increased cellular malleability and invasive behavior of the cancer cells.

A series of tasks were undertaken to accomplish the 3 aims in this proposal with predetermined milestones, and the results are described under each milestone.

**Aim 1.** We will determine/confirm if nesprin-1 expression is commonly lost in malignant breast cancer tissues and cell lines, compared to and correlated with benign tumors and primary human breast cancer cells.

**Aim 2.** We will determine if suppression (by siRNA and shRNA) of nesprin-1 in primary breast epithelial cells and non-metastatic breast cancer cells affects cell growth, leads to nuclear morphological deformation, and increases mobility and metastatic potential.

**Aim 3.** We will test if restoration of nesprin-1 expression (by cDNA transfection) in malignant breast cancer cells affects cell growth and suppress mobility and metastatic potential.

Task 1 (month 1-2): Collect 15 breast cancer cell lines and 3 non-cancer primary human mammary epithelial cell lines. Grow and expand these cells in cell culture, isolate cell lysate and mRNA for further analysis.

Task 2 (month 2): Use the mRNA and cell lysates prepared to perform qRT-PCR and Western blot to measure expression of nesprin-1. Multiple antibodies against the C- and N-terminal portion of nesprin-1 will be used to detect several spliced and alternative promoter isoforms. We will find out if nesprin-1 is commonly lost in breast cancer cells compared to non-cancer mammary epithelial cells, and if the loss of nesprin-1 correlates with malignancy of the cells.

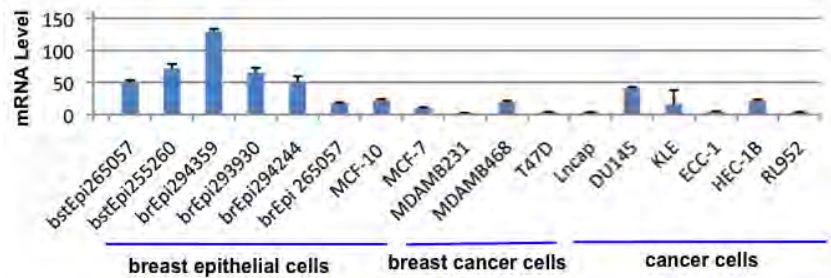
Task 3 (month 3): Perform qRT-PCR for nesprin-1 in total RNA isolated from 50 frozen breast cancers, with at the least 3 non-cancer controls. We will be able to determine if loss of nesprin-1 correlates with tumor grades and degree of malignancy in breast cancer tissues.

Task 4 (month 3): Collect around 100 breast cancer samples in tissue microarray. Perform immunostaining for nesprin-1. We will be able to determine if loss of nesprin-1 correlates with invasion and metastasis in breast cancer tissues.

**Milestone 1:** Tasks 1-4 (Aim 1) will enable us to verify if nesprin-1 is commonly lost in breast cancer cells, and if the loss of nesprin-1 correlates with invasion and metastasis in breast cancer.

We speculated and planned to determine/confirm if nesprin-1 expression is commonly lost in malignant breast cancer tissues and cell lines, compared to and correlated with benign tumors and primary human breast cancer cells. Indeed, we found that nesprin-1 expression is commonly lost in malignant breast cancer tissues and cell lines (**Figure 1**). We used a new technology, the nano-string approach (6) to determine the expression of nesprin-1 mRNA in breast cancer cells comparing to non-cancer cells, and found that nesprin-1 mRNA level is low or absent in all breast cancer cells analyzed, and a much higher level of expression is present in non-cancer human mammary epithelial cells.

**Figure 1. Loss of the expression of Nesprin-1 (SYNE-1) in breast cancer cells.** The mRNA expression of nesprin1 was determined by nanoString technology (6) in cells. The probe for nesprin-1 locates in the c-terminal KASH domain of human nesprin-1 gene. Five lines of primary cultures of human breast epithelial cells, a non-tumorigenic breast epithelial cell line (MCF-10), 4 breast cancer cell lines (MCF-7, MDAMB231, MDAMB468, and T47D), and 6 additional prostatic and endometrial cancer cell lines were analyzed. Generally, nesprin-1 expression is absent or greatly reduced in most cancer lines comparing to the non-tumorigenic breast epithelial cell line.



We purchased and tested 4 commercially available antibodies to nesprin-1 for Western blot and immunostaining. However, we have not been able to demonstrate that the antibodies recognized a nesprin-1 protein species or provided true staining in immunofluorescence microscopy. We concluded that these antibodies are not of sufficiently good quality for our experiments. We are currently in the process of making several monoclonal antibodies to peptides of human and mouse nesprin-1 sequences. Although these antibodies will not be available yet, they will be useful for future studies based on this pilot project. These tasks have enabled us to conclude that nesprin-1 expression is lost or greatly reduced in breast and other cancer.

Task 5 (month 4-6): siRNA will be prepared to target nesprin-1, and at the least two effective sequences will be used. A panel of breast cancer cells (MCF-7, T47D) identified in Aim 1 that have high nesprin-1 expression, will be transfected with siRNA to suppress nesprin-1 expression. Three lines of primary human mammary epithelial cells will be also tested. The transfected cells will be analyzed for suppression of nesprin-1 expression by Western blot.

Task 6 (month 6): The siRNA transfected cells will be analyzed for nuclear morphology by immunofluorescence microscopy, to observe the potential deformed cancer cell nuclear morphology following nesprin-1 suppression. The cells will be analyzed (comparing to scrambled siRNA controls) for growth, mobility, Boyden chamber invasion, and xenograft in nude mice to determine tumorigenesis and metastasis.

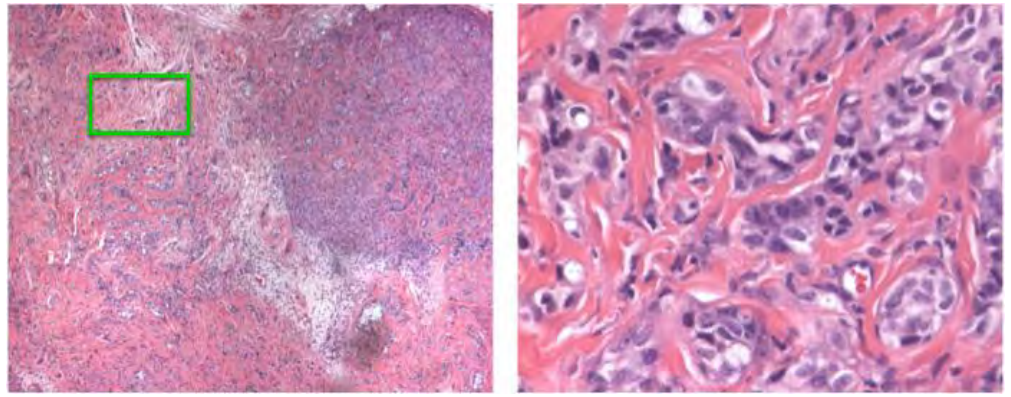
**Milestone 2:** These experiments in Task 5 and 6 (Aim 2) will determine/confirm if down regulation of nesprin-1 is sufficient to induce deformed nuclear morphology and increased mobility/invasion of the non-cancer mammary epithelial cells and non-metastatic cancer cells.

From the experimental results in **Aim 1**, we determined that the majority of breast cancer cell lines have low nesprin-1 expression, thus, no suitable cancer cell lines can be used for suppression of nesprin-1. When the expression of nesprin-1 was suppressed by siRNA and shRNA in primary breast epithelial cells (**Aim 2**), we found that the suppression led to nuclear morphological deformation, though we have not been able to sufficiently analyze cell growth and metastatic potential due to technical reason. The suppression of nesprin-1 is not sufficiently effective and long lasting for proper measurement of cell growth and mobility. The relatively short lifespan of primary human breast epithelial cells makes such experiments difficult.

In order to test the idea in this aim and to reach milestone, we used an available ovarian surface epithelial cell preparation available to us in the lab. The cells were prepared from a p53 null mouse and can grow well in tissue culture for extensive period. We transfected siRNA vector to target nesprin-1 sequence. The cells were implanted into nude mice to test tumorigenesis and tumor histology. In 3 out of 4 nude mice injected, tumors developed following 4 months, indicating a prolong latency. The tumors in all three mice exhibit similar histology: tumor cells are distributed in small nodules and intermingled with muscle fibers and stroma (**Figure 2**). The histology suggests that nesprin-1 suppressed cells are highly mobile and invasive. Thus, basically, the preliminary data is supportive of the hypothesis that loss of nesprin-1 may promote tumor cell mobility and invasion. The experiments enable us reaching **Milestone 2**.

**Figure 2. Loss of Nesprin-1 (SYNE-1) expression promotes tumor cell mobility and invasiveness.**

The representative H&E images of tumor developed in nude mice from nesprin-1 suppressed cells are shown in lower magnification (left panel) and high magnification (right panel, of the boxed area).



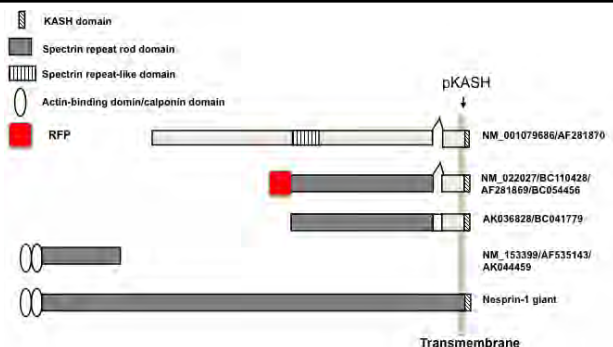
Task 7 (month 4-8): In 3 lines of nesprin-1-negative (determined in Aim 1) metastatic breast cancer cells, nesprin-1 will be transfected and expressed. The transfected clones will be selected by neo-resistance and the clones will be expanded.

Task 8 (month 9-11): The nesprin-1 expressing cells will be assayed for growth, mobility, invasion, and tumorigenesis/metastasis in nude mice, comparing to parental lines.

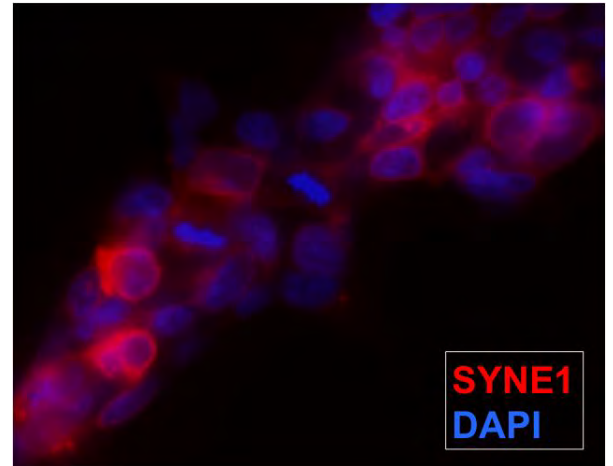
**Milestone 3:** Task 7-8 will accomplish Aim 3: To verify if nesprin-1 is a metastatic suppressor gene.

We also tested if restoration of nesprin-1 expression (by cDNA transfection) in malignant breast cancer cells affects cell growth and suppress mobility and metastatic potential (**Aim 3**). Since we are not able to monitor the expression of nesprin-1 definitely because of lacking a suitable antibody, we used a mCherry fusion to visualize nesprin-1 expression. Nesprin-1 gene produces multiple protein isoforms (**Figure 3**), we tested the 125 kd KASH domain containing isoform in our study. Expression of mCherry-nesprin-1 in T47D cells shows localization of the protein as a ring around nucleus as well as distribute in cytoplasm (**Figure 4**). Re-expression of mCherry-nesprin-1 was found to alter nuclear envelope morphology. However, the expressed nesprin-1 is not sufficiently stable to produce other significant phenotypes in the transfected cells.

**Figure 3. The structural isoforms of Nesprin-1/Syne1. illustrated.** The several known SYNE1/nesprin-1 protein isoforms derived from the gene are illustrated to indicate the actin binding calponin domain, KASH domain, and the Spectrin repeat rod domain (3). The 120 kd isoform of nesprin-1 was fused with mCherry and used for transfection into breast cancer cells.



**Figure 4. Lamin A expression in T47D breast cancer cells: immunofluorescence microscopy.** T47D breast cancer cells were transfected and selected for mCherry-nesprin-1 expression. The expression of mCherry-nesprin-1 (SYNE1) (red) was visualized by fluorescence microscope. DAPI staining (blue) was used to visualize nucleus.



Task 10 (month 12-14): Prepare final report to DOD. Prepare manuscript to report the potential findings!

**Milestone 4:** We will be able to conclude whether nesprin-1 is a breast cancer metastatic suppressor gene, and whether loss of nesprin-1 accounts for both nuclear deformation and cellular malleability of malignant breast cancer cells.

The results of these pilot experiments support the initial hypothesis of nesprin-1 as a metastatic suppressor gene and as an underlying link between two prominent features of a malignant cell, nuclear deformation and cellular malleability. The **Milestone 4**, obtaining evidence to support a metastasis suppressor role of nesprin-1, has been basically achieved. We also realize further complexity of nesprin-1 function in breast cancer suppression. The pilot study provides basis and promotes us to seek further investigation into the role of nesprin-1 in cancer malignancy.

#### KEY RESEARCH ACCOMPLISHMENTS:

This research project has been completed to satisfactory and enabled us to obtain supportive information for the overall hypothesis that nesprin-1 is lost in breast cancer and this may promote tumor cell invasion. The results are preliminary, though we are able to reach the following 3 conclusions.

1. We found that nesprin-1 expression is commonly lost in malignant breast cancer tissues and cell lines.
2. When the expression of nesprin-1 was suppressed by siRNA and shRNA in primary breast epithelial cells, we found that the suppression led to nuclear morphological deformation.
3. We found that suppression of nesprin-1 expression does not significantly influence cell proliferation, but enhanced metastasis.

#### REPORTABLE OUTCOMES:

The concept award allowed us to make initial study of our hypothesis. The preliminary results support the hypothesis that a defective nuclear envelope structure, due to the loss of nuclear envelope structure protein nesprin-1/SYNE-1, may be the underlying mechanism for nuclear morphological deformation and mobility and invasiveness of malignant breast cancer cells. The pilot results will be used to support application of additional fund to continue

## **CONCLUSION:**

The results of these pilot experiments support the initial hypothesis of nesprin-1 as a metastatic suppressor gene and as an underlying link between two prominent features of a malignant cell, nuclear deformation and cellular malleability. We also realize further complexity of nesprin-1 function in breast cancer suppression. The pilot study promotes us to seek further investigation into the role of nesprin-1 in cancer malignancy.

## **REFERENCES:**

1. Pienta KJ, Coffey DS. Correlation of nuclear morphometry with progression of breast cancer. *Cancer* 1991; 68:2012-2016.
2. Zink D, Fischer AH, Nickerson JA. Nuclear structure in cancer cells. *Nat Rev Cancer* 2004; 4:677-687.
3. Warren DT, Zhang Q, Weissberg PL, Shanahan CM. Nesprins: intracellular scaffolds that maintain cell architecture and coordinate cell function? *Expert Rev Mol Med* 2005;7(11):1-15.
4. Crisp M, Liu Q, Roux K, Rattner JB, Shanahan C, Burke B, Stahl PD, Hodzic D. Coupling of the nucleus and cytoplasm: role of the LINC complex. *J Cell Biol* 2006;172(1):41-53.
5. Olins AL, Hoang TV, Zwerger M, Herrmann H, Zentgraf H, Noegel AA, Karakesisoglou I, Hodzic D, Olins DE. The LINC-less granulocyte nucleus. *Eur J Cell Biol* 2009;88:203-214.
6. Geiss GK, Bumgarner RE, Birditt B, Dahl T, Dowidar N, Dunaway DL, Fell HP, Ferree S, George RD, Grogan T, et al.: Direct multiplexed measurement of gene expression with color-coded probe pairs. *Nat Biotechnol* 2008, 26:317-325.

## **APPENDICES:**

**None**

## **SUPPORTING DATA:**

**None**